



INTRODUCTION

It was recently shown that key biological control systems (e.g. MAPK pathway) are highly sensitive to the frequency of external stimuli in a non-intuitive manner which cannot be predicted by conventional pharmacometrics approaches [1]. This sug-

gests that quantitative systems pharmacology (QSP) can provide novel insights into optimal dosing regimens which could add a new dimension to the design of novel treatments. However, methods for such an approach are currently lacking. Recently, we illustrat-

ed the utility of frequency-domain response analysis (FdRA), a method widely used engineering, using several generic PK/PD case studies [2]. We now demonstrate the use of FdRA to optimise treatment regimen for cell-cycle specific chemotherapy.

CONCLUSION

Frequency-domain response analysis...

... identifies drug dosing frequencies for which the plasma concentration amplitude is attenuated/amplified in the response.

... identifies effective and save drug dosing frequencies for the cell-cycle specific tumour growth model.

... suggest metronomic chemotherapy as optimal cancer treatment.

METHODS

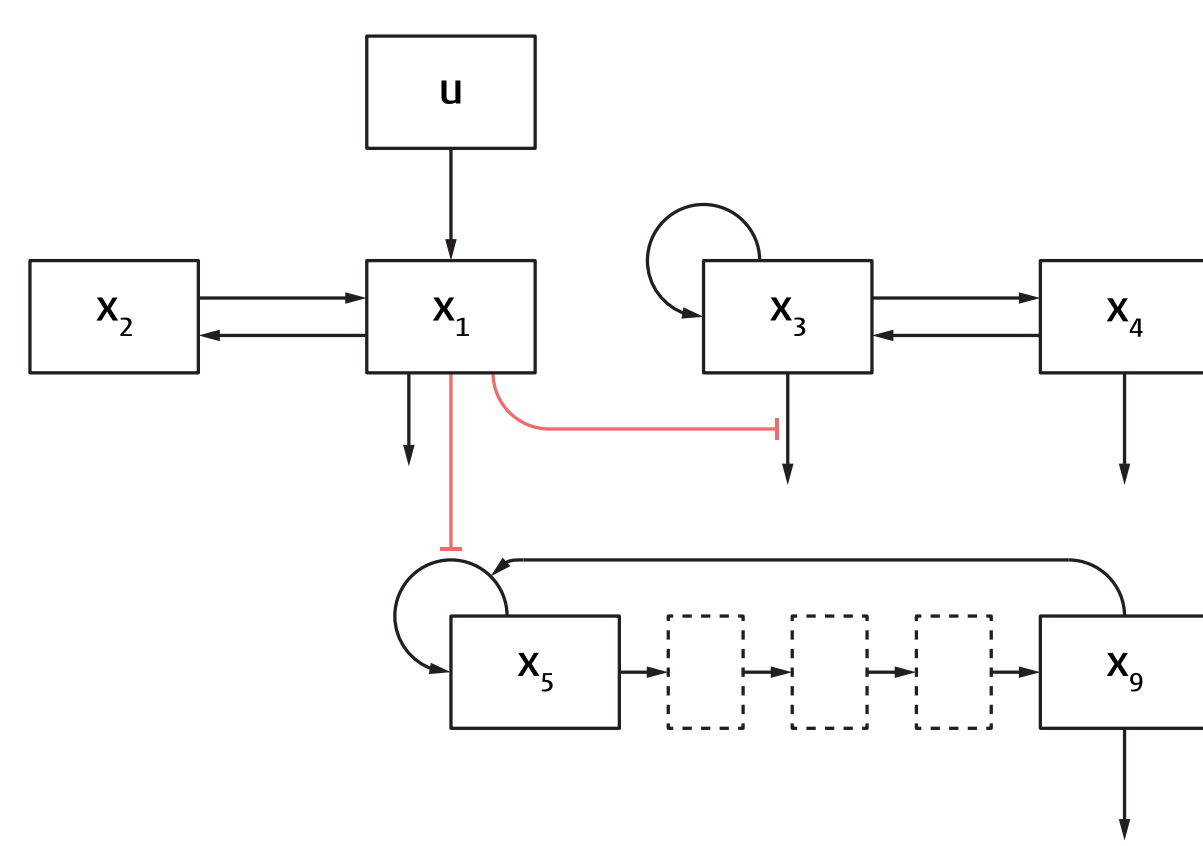


Figure 1: Two-comp. PK of Etoposide (x_1, x_2) linked to cell-cycle specific cancer model (x_3, x_4) and Friberg's model of myelosuppression (x_5 to x_9).

Analytically, frequency-domain response analysis (FdRA) determines the input/output behaviour of a linear, time-invariant (LTI) system

$$\dot{\mathbf{x}} = \mathbf{A}\mathbf{x} + \mathbf{b}u$$

$$\mathbf{y} = \mathbf{c}^T\mathbf{x} + \mathbf{d}u$$

in response to sinusoidal inputs of varying frequency. Thus, for the LTI system with \mathbf{x} , \mathbf{y} , and u denoting the model states, the outputs, and the inputs, respectively, and with $\mathbf{x}(0) = \mathbf{x}_0$ a so-called transfer function can be defined as

$$G(s) = \frac{Y(s)}{U(s)}$$

$$= \mathbf{c}^T (s\mathbf{I} - \mathbf{A})^{-1} \mathbf{b} + \mathbf{d}$$

The transfer function can now be used to collect the responses of the LTI system to sinusoidal inputs over a wide range of frequencies $s = i\omega$ with the help of a Bode plot. Its magnitude is defined as:

$$M(\omega) = \log_{10}|G(i\omega)|$$

and represents the logarithmic ratio of the output and the input amplitude.

Numerically, the frequency response can be determined for non-sinusoidal inputs that arise from repetitive dosing regimen commonly used in QSP modelling [2]. We here apply FdRA to a PK-

linked cell-cycle specific tumour growth model while considering safety by a model of myelosuppression (Figure 1).

The two-compartment PK model of Etoposide [3] is given by:

$$\dot{x}_1 = k_{21}x_2 - (k_{12} + k_e)x_1$$

$$\dot{x}_2 = k_{12}x_1 - k_{21}x_2$$

wherein x_1 and x_2 describe the plasma and peripheral compartment, respectively. Tumour growth dynamics are modeled by a cell-cycle specific two compartment model [4] in which proliferating cancer cells in G_1, S, G_2 , and M phase of the cell cycle are given by x_3 while quiescent cancer cells in G_0 phase are denoted by x_4

$$\dot{x}_3 = (\alpha - \mu - \eta)x_3 + \beta x_4 - \frac{k_1}{V_1}x_1x_3$$

$$\dot{x}_4 = \mu x_3 - (\beta + \gamma)x_4$$

Friberg's model of myelosuppression [5] describes stem and progenitor cells (x_5), three transit compartments of maturing cells (x_6, x_7 , and x_8), and circulating neutrophils (x_9) by:

$$\dot{x}_5 = k_7 x_5 \left(\delta(x_1) \frac{x_{90}^\lambda}{x_9^\lambda} - 1 \right)$$

$$\dot{x}_6 = k_7(x_5 - x_6)$$

$$\dot{x}_7 = k_7(x_6 - x_7)$$

$$\dot{x}_8 = k_7(x_7 - x_8)$$

$$\dot{x}_9 = k_7(x_8 - x_9)$$

RESULTS

The cell-cycle specific tumour growth model is now stimulated according to different dosing schemes. Exemplified in Figure 2, we assume constant exposure to Etoposide at the maximum tolerated dose of 500 mg/m² either at a dosing frequency of one dose every 60 days (blue lines) or at a dosing frequency of one dose every 7 days (green lines).

We observe that even though higher doses are administered every 60 days the tumour continues to grow while low doses every 7 days lead to overall decrease in tumour mass. Additionally, more frequent lower doses do not disrupt neutrophil count as strongly as less frequent higher doses.

In order to systematically study the response of cancer cells and neutrophils to different dosing schemes, we applied numerical FdRA to the model given in Figure 1. To this end, we administered doses at frequencies between one bolus dose every 120 days and 24 bolus doses per day

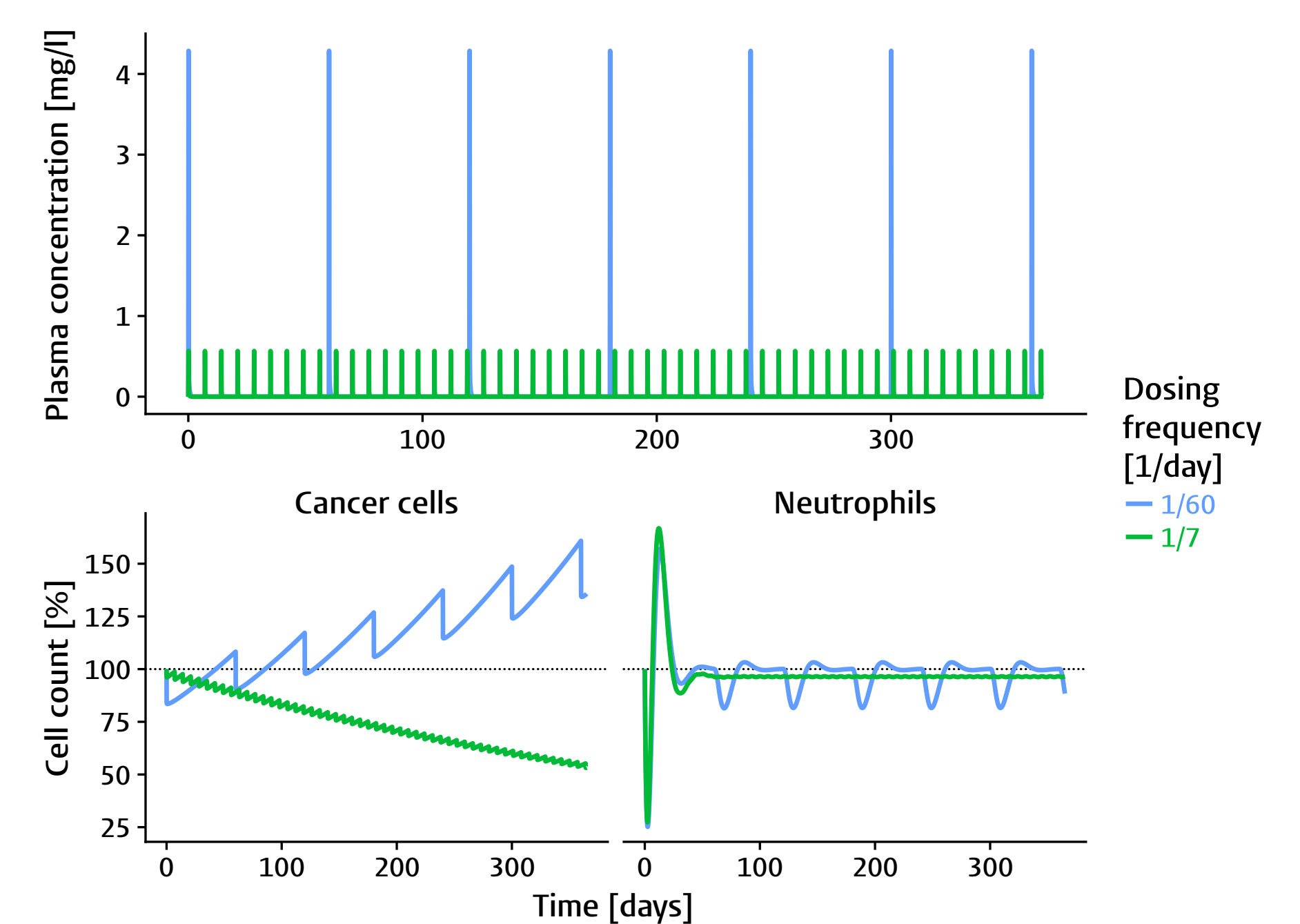


Figure 2: Cancer cell and neutrophil time courses for two dosing frequencies at constant exposure showing effective (green) and ineffective (blue) dosing.

at either variable (i.e. each dose equals 500 mg/m²) or constant exposure. Afterwards, we measured the amplitude of the plasma concentration of Etoposide as well as the amplitudes of the total number of cancer cells (proliferating + quiescent cells) and neutrophils after one year of treatment. The output to input amplitude ratio is then plotted over the dosing frequency in Figure 3.

We observe that only constant exposure negatively affects safety (i.e. neutrophil counts below 0.5 · 10⁹). Furthermore, less frequent dosing is ineffective and typically leads to a higher amplitude ratio. Only the neutrophil amplitude ratio under variable exposure increases for decreasing dosing frequencies while in all other cases the amplitude ratio decreases.

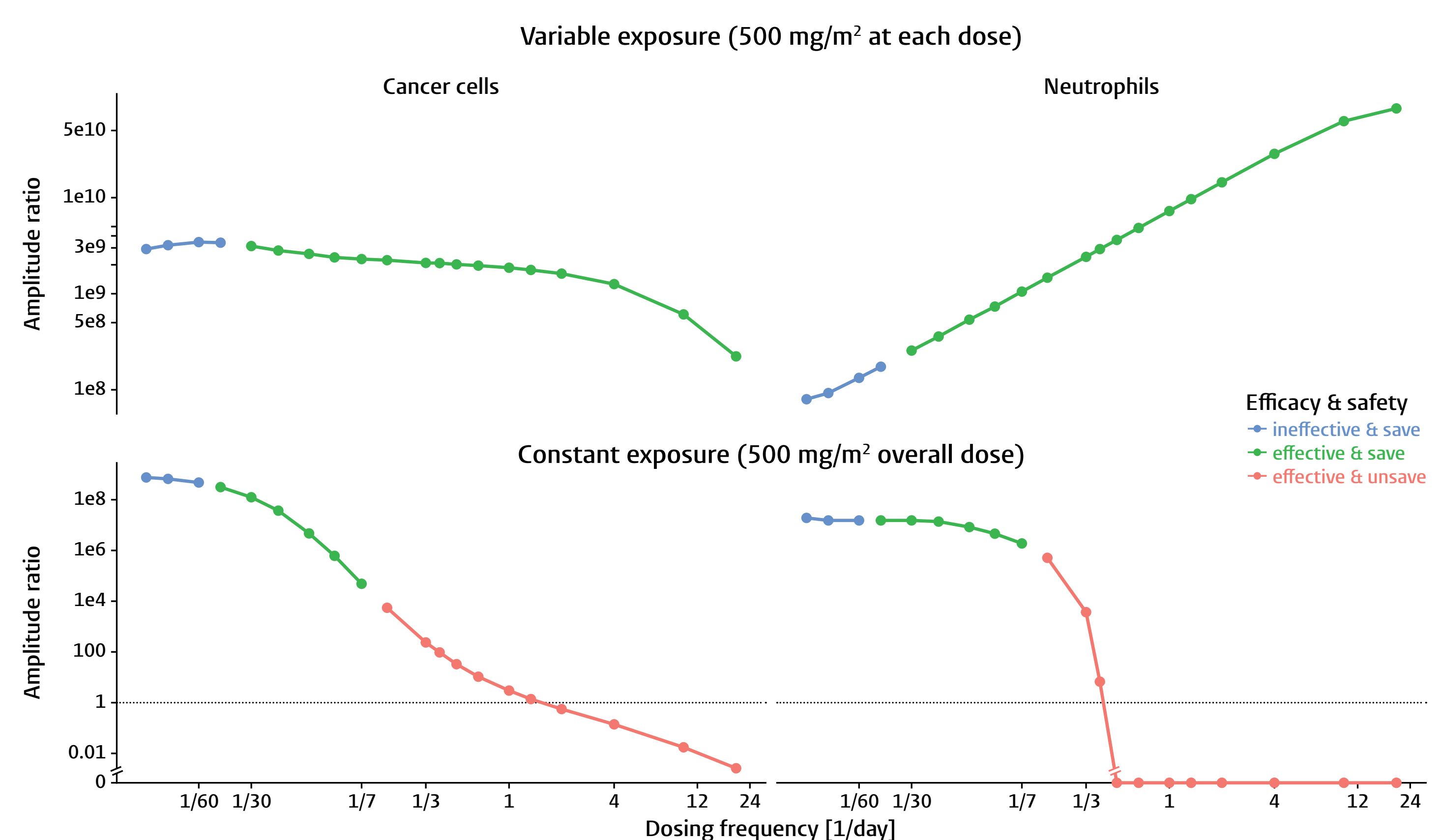


Figure 3: Cancer cell and neutrophil frequency response for constant and variable exposure.

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