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## Thermodynamic modeling explains the regulation of *CYP1A1* expression in the liver Thesis defense

**Pascal Schulthess** 





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## Exposure to dioxin through food uptake



welt.de, wikipedia.org, beyondthedish.wordpress.com

## CYPs biotransform toxins



### Cytochrome P450s (CYPs)

- major enzymes in detoxification
- harbor broad substrate specificity
- increase hydrophilicity of compounds to ease their excretion in bile or urine





Braeuning, A. & Schwarz, M. Biol. Chem. 391, 139–148 (2010) Braeuning, A., Köhle, C., Buchmann, A. & Schwarz, M. Toxicol. Sci. 122, 16–25 (2011).





#### CYP1A



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### How are the two signals integrated?





### How are the two signals integrated?

### How do binding sites <u>cooperate</u>?



### 1. Introduction of the modeling framework



2. Model of synthetic promoter



3. Model of wild-type CYP1A1 promoter





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## Modeling approaches for gene expression



Level of detail



## Mathematical model combines signaling and promoter logic



### Thermodynamic model

Signaling model





$$p_{\text{bound}}([P$$

 $) = \frac{\text{transcribing states}}{\text{all possible states}}$ 







 $) = \frac{\text{transcribing states}}{\text{all possible states}}$  $= \frac{q_P}{1 + q_P}$ 

with  $q_i = K_i[i]$ 





 $p_{\text{bound}}([P], [A])$ 

 $) = \frac{\text{transcribing states}}{\text{all possible states}}$ 

$$\frac{q_P + q_P q_A E_{PA}}{\Xi}$$

$$1 + q_P + q_A + q_P q_A E_{PA}$$

with 
$$q_i = K_i[i]$$
, and  $E_{ij} = \exp(-\epsilon_{ij}/k_BT)$ 





 $p_{\text{bound}}([P], [A], [B]) = \frac{\text{transcribing states}}{\text{all possible states}}$  $= \frac{q_P + q_P q_A E_{PA} + q_P q_B E_{PB} + q_P q_A q_B E_{PA} E_{PB} E_{AB}}{1 + q_P + q_A + q_P q_A E_{PA} + q_B + q_P q_B E_{PB} + q_A q_B E_{AB} + q_P q_A q_B E_{PA} E_{PB} E_{AB}}$ 

with 
$$q_i = K_i[i]$$
, and  $E_{ij} = \exp(-\epsilon_{ij}/k_BT)$ 



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 $1 \times C$   $2 \times C$   $3 \times C$   $4 \times C$   $5 \times C$   $6 \times C$ 

1× D 2× D









| model        | parameters |
|--------------|------------|
| signaling    | 3          |
| C constructs | 13 (28)    |
| D constructs | 1 (9)      |



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|--------------|------------|
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#### Only first TF interacts with polymerase





Only first TF interacts with polymerase

Only nearby TFs cooperate





**Only first TF interacts with polymerase** 

Only nearby TFs cooperate



#### **Only first TF interacts with polymerase**

### Only nearby TFs cooperate



mean number of occupied binding sites



mean number of occupied binding sites



probability that first binding site is occupied





mean number of occupied binding sites



probability that first binding site is occupied





mean number of occupied binding sites



probability that first binding site is occupied





mean number of occupied binding sites



probability that first binding site is occupied





mean number of occupied binding sites



probability that first binding site is occupied





mean number of occupied binding sites



probability that first binding site is occupied





### Sequestration responsible for reduced induction













Integration of the two signals follows AND gate logic



#### How do binding sites <u>cooperate</u>?

## Conclusion for synthetic promoter

### How do binding sites <u>cooperate</u>?

- only first binding site interacts with polymerase
- only nearby binding sites cooperate

## Conclusion for synthetic promoter

### How do binding sites <u>cooperate</u>?

- only first binding site interacts with polymerase
- only nearby binding sites cooperate

How are the two signaling pathways integrated?integration follows an AND gate logic



#### Does the same hold true for the

wild-type CYP1A1 promoter?



### 1. Introduction of the modeling framework



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Dioxin [nM]

| model         | parameters |
|---------------|------------|
| signaling     | 3          |
| thermodynamic | 16 (21)    |



Dioxin [nM]

| model         | parameters |
|---------------|------------|
| signaling     | 3          |
| thermodynamic | 16 (21)    |



synthetic promoter









### More and stronger TF-RNAP interactions



#### More and stronger TF-RNAP interactions

### **β-catenin is important interaction partner**





Dioxin [nM]











### **Gradual AND gate enables finer regulation**



#### How do binding sites <u>cooperate</u>?

## Conclusion for wild-type CYP1A1 promoter

### How do binding sites <u>cooperate</u>?

- more complex interaction patterns
- increased importance of TF-RNAP interactions
- β-catenin is an important interaction partner

## Conclusion for wild-type CYP1A1 promoter

### How do binding sites <u>cooperate</u>?

- more complex interaction patterns
- increased importance of TF-RNAP interactions
- β-catenin is an important interaction partner

- integration follows an AND gate logic
- gradual signal integration enables a finer regulation of promoter response

## Model qualitatively predicts hepatic zonation



## Model qualitatively predicts hepatic zonation







The interactions at the **signaling** level, together with the TF **cooperativity** in the *CYP1A1* promoter enable the **spatial expression** pattern observed *in vivo*.

Schulthess P, et al. (2015) Signal integration by the *CYP1A1* promoter - a quantitative study. *Nucleic Acids Research* 43(11):5318–5330.



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COMPUTATIONAL MODELLING IN MEDICINE









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